

APPLICATION

Fast Analysis of Sucrose, Glucose, and Fructose Composition in Fruit Juices and Processed Beverages using Simplified HPLC Methodology

Laura Snow, Matthew Trass, Brian Rivera, Michael Klein, and Allen Misa
Phenomenex, 411 Madrid Avenue, Torrance, CA 90501 USA



Brian Rivera
Product Manager

In addition to chromatography, Brian also has a passion for ice cream-making, and enjoys experimenting with bold, new flavors.

Sucrose, glucose, and fructose are naturally found in fruit juices but are also added to various processed beverages. In this study, we present a fast and simple method for the analysis of sugar using Rezex HPLC columns evaluating a variety of commonly consumed beverages including sports/energy drinks, soda, fruit juices, and wine.

Introduction

The quantitation of sucrose and its monosaccharide constituents, glucose and fructose, is of critical importance for beverage manufacturers. High-fructose corn syrup (HFCS) is a sweetener composed primarily of glucose and fructose, which is ubiquitous in processed foods and beverages. There is a concern for HFCS content in processed foods because HFCS has been linked to metabolic disorders, including diabetes¹. There is also a need for quantitation of sugar in naturally sweetened beverages since sucrose may degrade during the manufacturing process².

A number of HPLC methods have been used to quantitate sucrose, glucose, and fructose including HILIC³ and Ligand Exchange⁴. However, HILIC methods will use high amounts of organic and analysis might require extensive method development. Previous Ligand Exchange methods provide linearity and limits of quantitation that are not practical for real-world applications.

In this study, we demonstrate the use of RezexTM RCM-Monosaccharide Ca⁺² to develop a quick and quantitative method under 15 minutes for the analysis of the three main sugars in various beverages and fruit juices: sucrose, glucose, and fructose.

Materials and Methods

Reagents and Chemicals

Sugar standards were purchased from Sigma Chemical (St. Louis, MO). Beverages were obtained from local grocers. A Sartorius[®] arium[®] comfort II was used for ASTM Type 1 ultrapure water. Nine standards were prepared in water at concentrations from 0.1 mg/mL to 50 mg/mL. Beverage samples were diluted 1:10 in ultrapure

water. Although not performed in this study, the use of a 0.45 µm PhenexTM-NY (Nylon) syringe filter prior to injection is encouraged for beverages with high particulate content (e.g. fruit juice with pulp).

Experimental Conditions

HPLC analysis was performed using an Agilent[®] 1260 LC system (Agilent Technologies, Palo Alto, CA, USA) with an upper pressure limit of 600 bar, equipped with a binary pump, autosampler, and Refractive Index (RI) detector. Method conditions are below.

| | |
|----------------------|---|
| Column: | Rezex RCM-Monosaccharide Ca ⁺² |
| Dimensions: | 100 x 7.8 mm |
| Part No.: | 00D-0130-K0 |
| Mobile Phase: | Water |
| Flow Rate: | 0.4 mL/min |
| Temperature: | 80 °C |
| Detector: | RI (Refractive Index) @ 35 °C |
| Sample: | Sucrose Glucose Fructose |



Results and Discussion

A nine-point standard curve was run for sucrose, glucose, and fructose from 0.1 mg/mL to 50 mg/mL (**Table 1**). Retention times for standards are shown in **Table 2**. Using peak areas, a linear regression line was drawn for each sugar respectively, with correlation coefficients greater than 0.99 for each (**Figures 1-3**).

Column: Rezex™ RCM-Monosaccharide Ca²⁺
Dimensions: 100 x 7.8 mm
Part No.: 00D-0130-K0
Mobile Phase: Water
Flow Rate: 0.4 mL/min
Temperature: 80 °C
Detector: RI (Refractive Index) @ 35 °C
Sample: Sucrose
 Glucose
 Fructose

Table 1. Standard concentrations

| Standards | Conc. (mg/mL) |
|-----------|---------------|
| 1 | 0.1 |
| 2 | 0.25 |
| 3 | 0.5 |
| 4 | 1 |
| 5 | 2.5 |
| 6 | 5 |
| 7 | 10 |
| 8 | 25 |
| 9 | 50 |

Figure 1. Standard curve for sucrose, 0.1-50 mg/mL

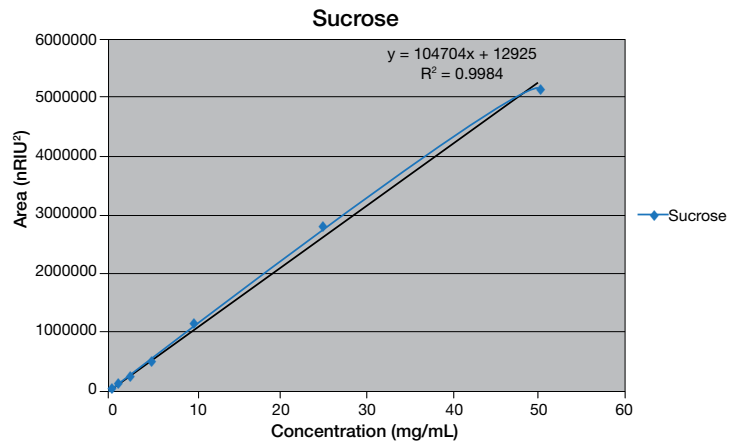


Figure 2. Standard curve for glucose, 0.1-50 mg/mL

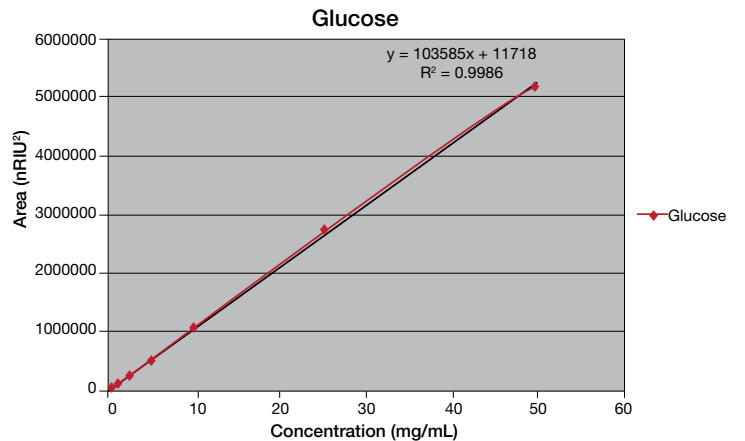
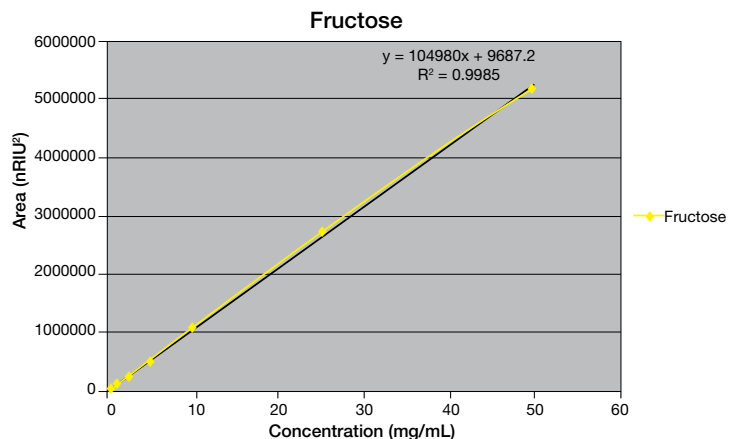


Figure 3. Standard curve for fructose, 0.1-50 mg/mL



Typically in ion exclusion and ligand exchange, a minimum of a one minute retention time difference is required for baseline separation. Baseline separation is clearly observed in **Figures 4 & 5**, at 0.1 mg/mL and 50 mg/mL, respectively.

Concentrations for each beverage were calculated using the calibration curve then multiplied by 10 to adjust for the dilution factor (**Table 3**).

Figure 4. 0.1 mg/mL Sugar Standard, prepared in Water

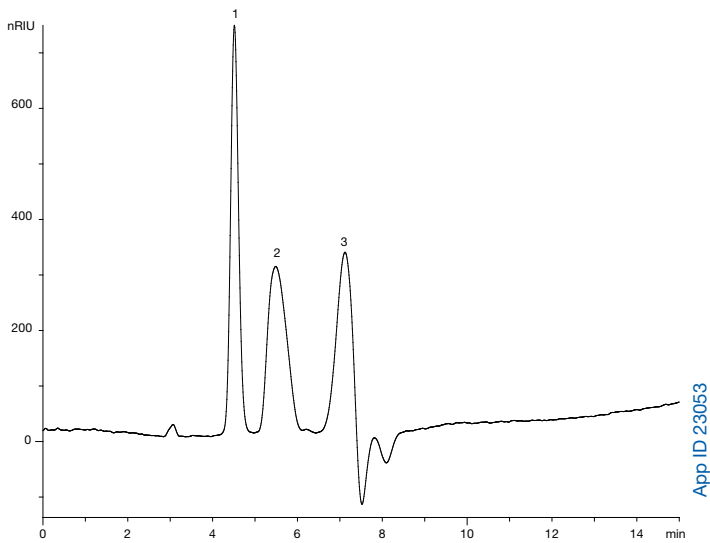
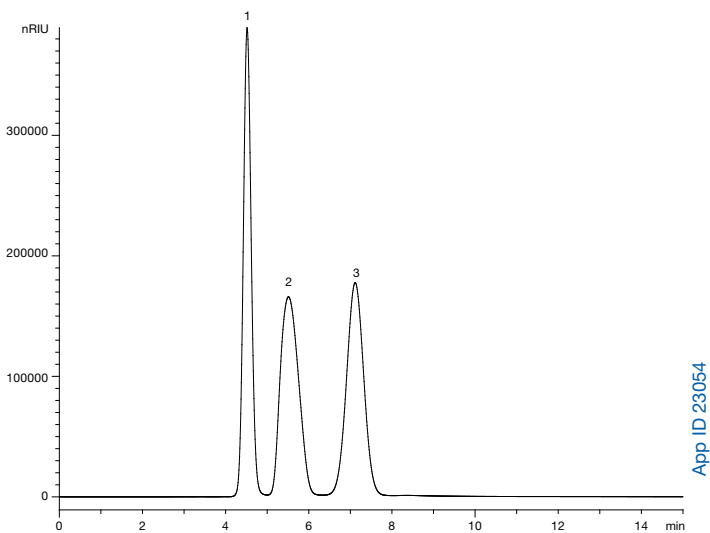


Table 2. Standard Retention Times

| Sugar | Retention Time (min) |
|-------------|----------------------|
| 1. Sucrose | 4.5 |
| 2. Glucose | 5.5 |
| 3. Fructose | 7.1 |

Figure 5. 50 mg/mL Sugar Standard, prepared in Water



Column: Rezex RCM-Monosaccharide Ca²⁺
Dimensions: 100 x 7.8 mm
Part No.: 00D-0130-K0
Mobile Phase: Water
Flow Rate: 0.4 mL/min
Temperature: 80 °C
Detector: RI (Refractive Index) @ 35 °C
Sample: Sucrose
 Glucose
 Fructose



Percent deviations were determined by comparing the concentration of the three sugars combined to expected concentration of sugars according to the nutrition facts on the beverage label. **Figures 6 & 7** show sugar content in red and white wine, respectively. Red wine in particular can have fructose content which gives an undesirable sweetness⁵. It has been reported that between 1-2.5% sugar content⁶ can be detected by most palates and the linearity of the assay (i.e. 0.1-5%) is appropriate for this analysis.

Table 3. Analytes and Sugar Content in Comparison to Reported Nutrition Facts

| Beverages | Sucrose (mg/mL) | Glucose (mg/mL) | Fructose (mg/mL) | Total Sugars (mg/mL) | Percent Deviation from Nutrition Facts |
|------------------------|-----------------|-----------------|------------------|----------------------|--|
| Merlot (Red Wine) | 0.0 | 0.0 | 1.6 | 2.1 | N/A |
| Moscato (White Wine) | 0.0 | 26.8 | 51.3 | 76.9 | N/A |
| Orange Juice 1 | 52.7 | 29.2 | 30.6 | 112.6 | 28.63% |
| Orange Juice 2 | 50.9 | 26.5 | 29.5 | 106.8 | 16.55% |
| Fruit Punch | 4.3 | 51.3 | 69.8 | 125.4 | 36.79% |
| Pineapple Orange Juice | 35.0 | 34.5 | 46.9 | 116.4 | 3.43% |
| Pineapple Mango Juice | 35.7 | 23.5 | 38.1 | 97.4 | 16.90% |
| Energy Drink | 40.4 | 54.7 | 30.6 | 125.8 | 10.16% |
| Cola | 2.5 | 49.8 | 66.7 | 119.0 | 1.92% |
| Sports Drink | 38.2 | 15.8 | 11.9 | 65.9 | 14.52% |

Figure 6. Merlot (Red Wine)

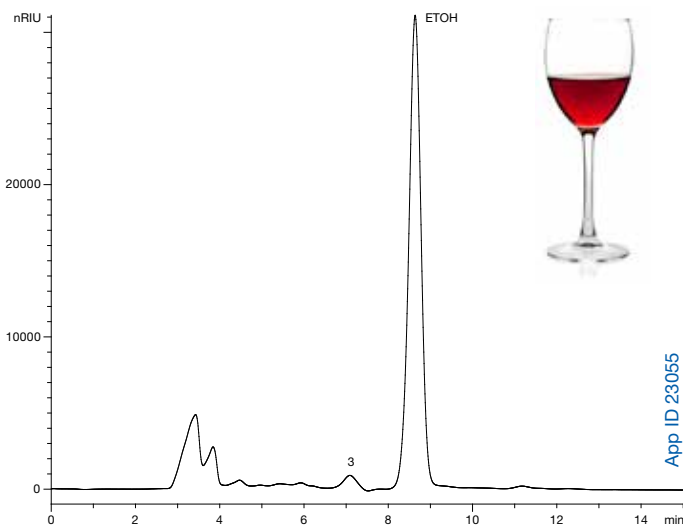
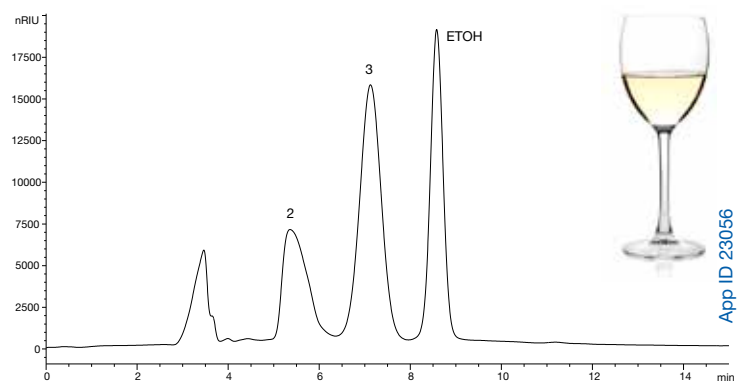


Figure 7. Moscato (White Wine)



Figures 8 & 9 show sugar content for orange juices from two separate sources. Both are similar in sugar content. Figure 10 shows an artificially flavored “fruit punch.” Only 4.3mg/mL sucrose is detected, indicating sweetening primarily with HFCS.

Figure 8. Orange Juice 1

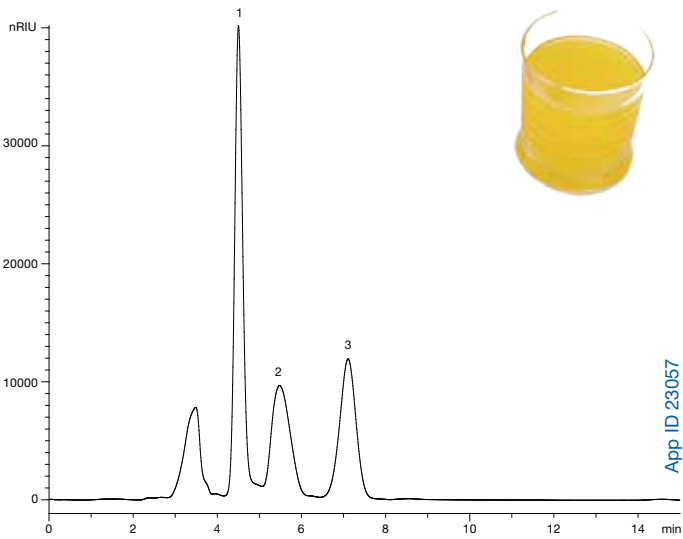


Figure 9. Orange Juice 2

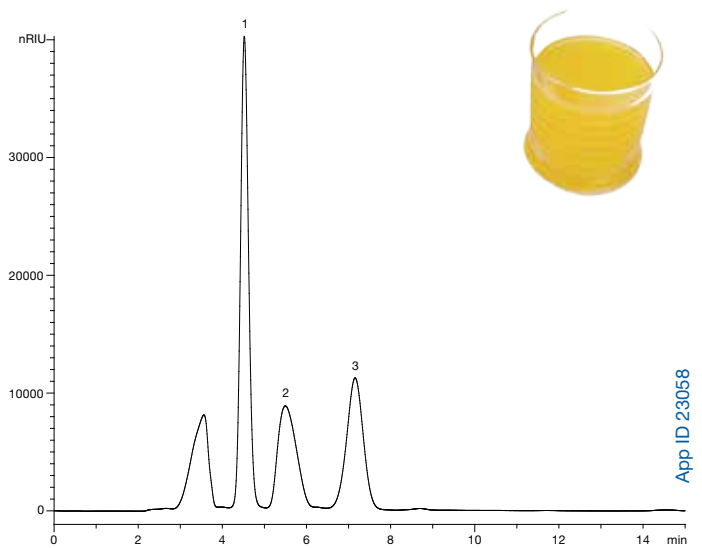
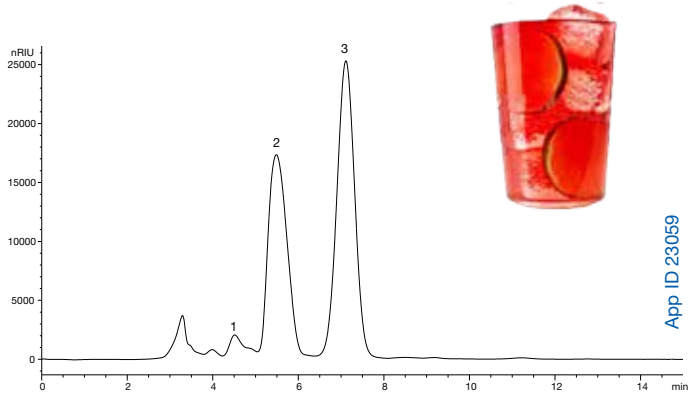


Figure 10. Fruit Punch



Column: Rezex RCM-Monosaccharide Ca²⁺
Dimensions: 100 x 7.8 mm
Part No.: 00D-0130-K0
Mobile Phase: Water
Flow Rate: 0.4 mL/min
Temperature: 80 °C
Detector: RI (Refractive Index) @ 35 °C
Sample: Sucrose
 Glucose
 Fructose



The two samples “Pineapple Orange Juice” and “Pineapple Mango Juice” are reported to be 100% juice. These juices can also contain the natural sugar alcohol sorbitol. Note in this method, the sorbitol peak elutes at approximately 12 minutes (**Figures 11 & 12**) Although not the focus of this study, this method could feasibly be used for sorbitol quantitation.

Figure 11. Pineapple Orange Juice

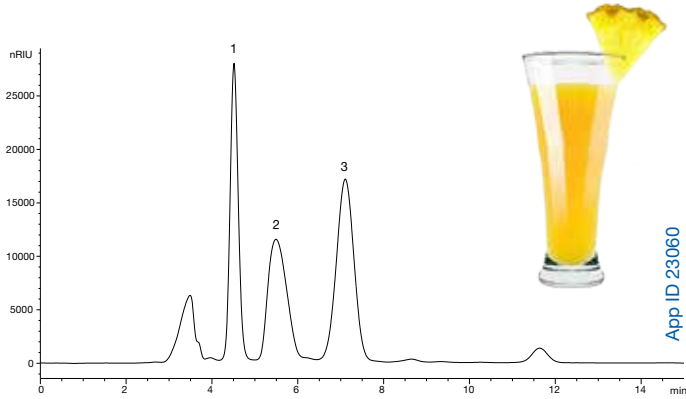
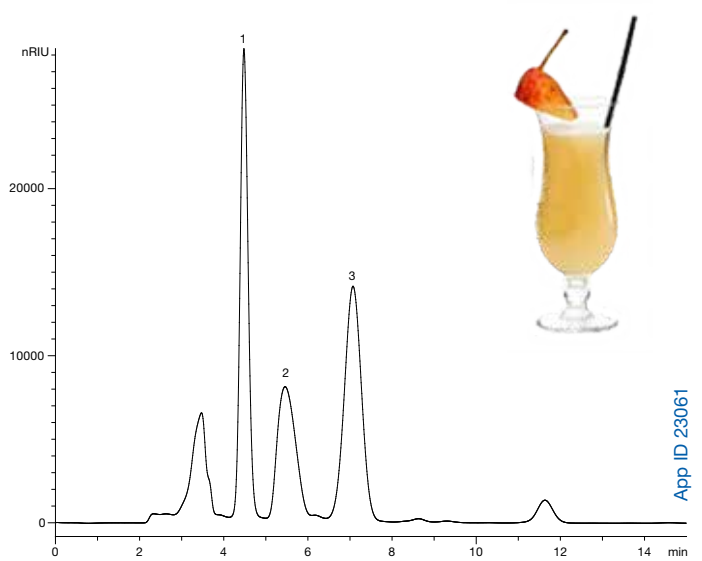


Figure 12. Pineapple Mango Juice



The “Energy Drink” sample contained two unknown peaks eluting at approximately 7-8 minutes (**Figure 13**). Further studies would need to be performed to confirm the identity of these peaks. Cola sample in **Figure 14** shows almost no sucrose, indicating the beverage is sweetened primarily with HFCS. Conversely, “Sports Drink” sample showed it primarily being sweetened with sucrose (**Figure 15**).

Figure 13. Energy Drink

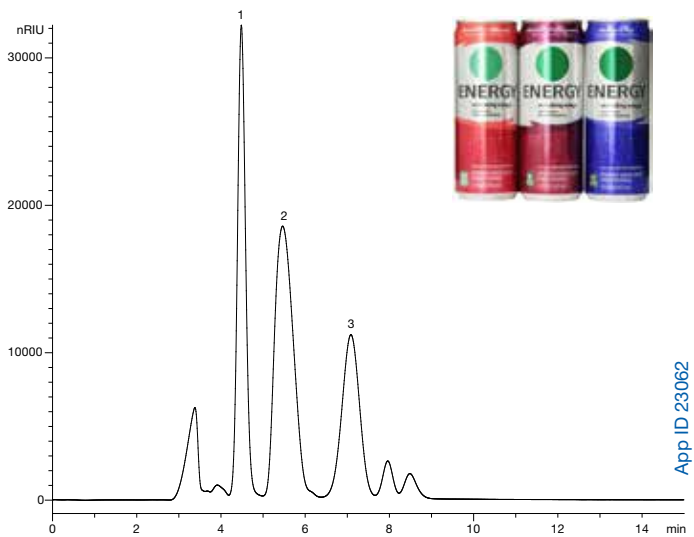


Figure 14. Cola

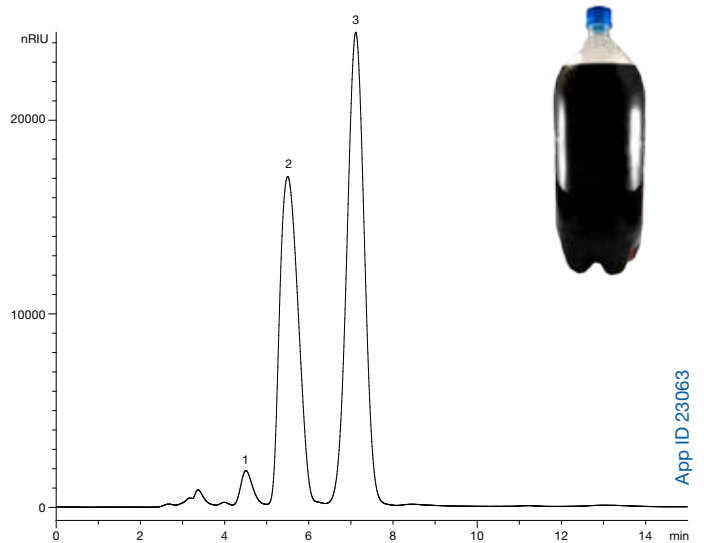
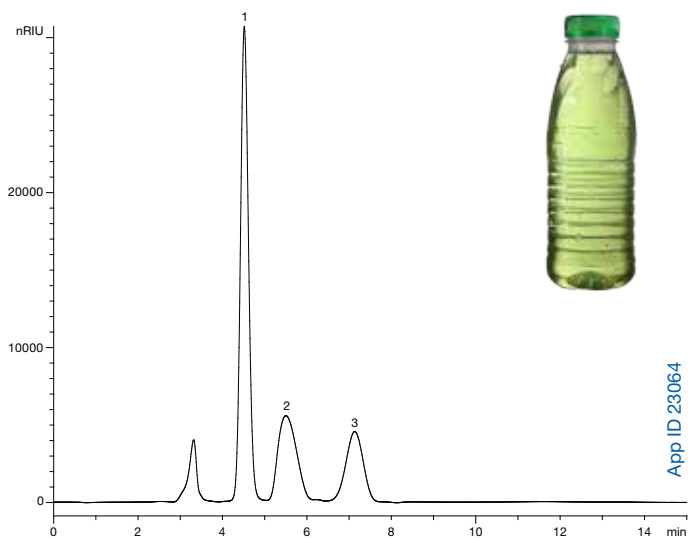


Figure 15. Sports Drink



Finally, it is worth noting that both “Orange Juice 1” and “Fruit Punch” samples were outside the 20% discrepancy allowed by the FDA⁷.

Conclusion

In this study, we demonstrated a method using Rezex™ RCM for the analysis of sucrose, glucose, and fructose content in processed sugary drinks and beverages. The method is quantitative, simple, and robust, being compatible with a variety of sample matrices, including wines, fruit juices, and sports drinks. Further studies could include the inclusion of sorbitol, as well as other sugar alcohols.

References

1. Chan, Amanda L. (2012-11-27). “Is There A Link Between High Fructose Corn Syrup And Diabetes Rates?”. Huffington Post.
2. Shahidi, Fereidoon, Chi-Tang Ho, and Nguyen Van Chuyen. Process-induced Chemical Changes in Food. Vol. 434. S.I.: Springer-Verlag New York, 2013. Print.
3. Ma, Chunmei, Zhen Sun, Changbao Chen, Lili Zhang, and Shuhua Zhu. “Simultaneous Separation and Determination of Fructose, Sorbitol, Glucose and Sucrose in Fruits by HPLC–ELSD.” Food Chemistry (2014): 784-88. Print.
4. Mahmood, Tahir et al. “Compositional Variation in Sugars and Organic Acids at Different Maturity Stages in Selected Small Fruits from Pakistan.” International Journal of Molecular Sciences 13.2 (2012): 1380–1392. PMC. Web. 17 May 2015.
5. “Sugars in Wine.” Wikipedia. Wikimedia Foundation. Web. 17 May 2015.
6. Berthels, N., R. Cordero, F. Bauer, J. Thevelein, and I. Pretorius. “Discrepancy in Glucose and Fructose Utilisation during Fermentation by Wine Yeast Strains.” FEMS Yeast Research (2004): 683-89. Print.
7. Bender, Mary M, Jeanne I. Rader, and Foster D. McClure. “Guidance for Industry: FDA Nutrition Labeling Manual—A Guide for Developing and Using Data Bases.” US Dept of Health and Human Services, Food and Drug Administration Center for Food Safety and Applied Nutrition. Web. 17 May 2015

Rezex Ordering Information

| Columns | | | | | Guards | | SecurityGuard™ Cartridges (mm) |
|------------------------|-------------|---------------|------------|------------|-------------|-----------|--------------------------------|
| Description | Part No. | Cross Linkage | Ionic Form | Size (mm) | Part No. | Size (mm) | 4 x 3.0* |
| RCM-Monosaccharide | 00D-0130-KO | 8% | Calcium | 100 x 7.8 | 03B-0130-KO | 50 x 7.8 | AJO-4493 |
| RCM-Monosaccharide | 00F-0130-KO | 8% | Calcium | 150 x 7.8 | 03B-0130-KO | 50 x 7.8 | AJO-4493 |
| RCM-Monosaccharide | 00H-0130-KO | 8% | Calcium | 300 x 7.8 | 03B-0130-KO | 50 x 7.8 | AJO-4493 |
| RHM-Monosaccharide | 00H-0132-KO | 8% | Hydrogen | 300 x 7.8 | 03B-0132-KO | 50 x 7.8 | AJO-4490 |
| RAM-Carbohydrate | 00H-0131-KO | 8% | Silver | 300 x 7.8 | — | — | AJO-4491 |
| RSO-Oligosaccharide | 00P-0133-NO | 4% | Silver | 200 x 10.0 | 03R-0133-NO | 60 x 10.0 | — |
| RNO-Oligosaccharide | 00P-0137-NO | 4% | Sodium | 200 x 10.0 | 03R-0137-NO | 60 x 10.0 | — |
| RPM-Monosaccharide | 00H-0135-KO | 8% | Lead | 300 x 7.8 | 03B-0135-KO | 50 x 7.8 | AJO-4492 |
| (for USP procedure) | 00D-0135-KO | 8% | Lead | 100 x 7.8 | 03B-0135-KO | 50 x 7.8 | AJO-4492 |
| RNM-Carbohydrate | 00H-0136-KO | 8% | Sodium | 300 x 7.8 | 03B-0136-KO | 50 x 7.8 | — |
| ROA-Organic Acid | 00F-0138-E0 | 8% | Hydrogen | 150 x 4.6 | — | — | AJO-4490 |
| ROA-Organic Acid | 00G-0138-E0 | 8% | Hydrogen | 250 x 4.6 | — | — | AJO-4490 |
| ROA-Organic Acid | 00F-0138-KO | 8% | Hydrogen | 150 x 7.8 | 03B-0138-KO | 50 x 7.8 | AJO-4490 |
| ROA-Organic Acid | 00H-0138-KO | 8% | Hydrogen | 300 x 7.8 | 03B-0138-KO | 50 x 7.8 | AJO-4490 |
| RKP-Potassium | 00H-3252-KO | 8% | Potassium | 300 x 7.8 | — | — | — |
| RFQ-Fast Acid | 00D-0223-KO | 8% | Hydrogen | 100 x 7.8 | 03B-0223-KO | 50 x 7.8 | AJO-4490 |
| RCU-USP Sugar Alcohols | 00G-0130-D0 | 8% | Calcium | 250 x 4.0 | — | — | AJO-4493 |

for ID: 3.2-8.0 mm

*SecurityGuard Analytical Cartridges require universal holder Part No.: KJO-4282



If Phenomenex products in this technical note do not provide at least equivalent separations as compared to other products of the same phase, size and dimensions, return the product with comparative data within 45 days for a FULL REFUND.

APPLICATION

Australia

t: +61 (0)2-9428-6444
 f: +61 (0)2-9428-6445
 auinfo@phenomenex.com

Austria

t: +43 (0)1-319-1301
 f: +43 (0)1-319-1300
 anfrage@phenomenex.com

Belgium

t: +32 (0)2 503 4015 (French)
 t: +32 (0)2 511 8666 (Dutch)
 f: +31 (0)30-2383749
 beinfo@phenomenex.com

Canada

t: +1 (800) 543-3681
 f: +1 (310) 328-7768
 info@phenomenex.com

China

t: +86 (0)20 2282-6668
 f: +86 (0)20 2809-8130
 chinainfo@phenomenex.com

Denmark

t: +45 4824 8048
 f: +45 4810 6265
 nordicinfo@phenomenex.com

Finland

t: +358 (0)9 4789 0063
 f: +45 4810 6265
 nordicinfo@phenomenex.com

France

t: +33 (0)1 30 09 21 10
 f: +33 (0)1 30 09 21 11
 franceinfo@phenomenex.com

Germany

t: +49 (0)6021-58830-0
 f: +49 (0)6021-58830-11
 anfrage@phenomenex.com

India

t: +91 (0)40-3012 2400
 f: +91 (0)40-3012 2411
 indiainfo@phenomenex.com

Ireland

t: +353 (0)1 247 5405
 f: +44 1625-501796
 eireinfo@phenomenex.com

Italy

t: +39 051 6327511
 f: +39 051 6327555
 italiainfo@phenomenex.com

Luxembourg

t: +31 (0)30-2418700
 f: +31 (0)30-2383749
 nlinfo@phenomenex.com

Mexico

t: 001-800-844-5226
 f: 001-310-328-7768
 tecnicomx@phenomenex.com

The Netherlands

t: +31 (0)30-2418700
 f: +31 (0)30-2383749
 nlinfo@phenomenex.com

New Zealand

t: +64 (0)9-4780951
 f: +64 (0)9-4780952
 nzinfo@phenomenex.com

Norway

t: +47 810 02 005
 f: +45 4810 6265
 nordicinfo@phenomenex.com

Puerto Rico

t: +1 (800) 541-HPLC
 f: +1 (310) 328-7768
 info@phenomenex.com

Spain

t: +34 91-413-8613
 f: +34 91-413-2290
 espinfo@phenomenex.com

Sweden

t: +46 (0)8 611 6950
 f: +45 4810 6265
 nordicinfo@phenomenex.com

United Kingdom

t: +44 (0)1625-501367
 f: +44 (0)1625-501796
 ukinfo@phenomenex.com

USA

t: +1 (310) 212-0555
 f: +1 (310) 328-7768
 info@phenomenex.com

**All other countries
Corporate Office USA** 

t: +1 (310) 212-0555
 f: +1 (310) 328-7768
 info@phenomenex.com

www.phenomenex.com

Phenomenex products are available worldwide. For the distributor in your country, contact Phenomenex USA, International Department at international@phenomenex.com

Terms and Conditions

Subject to Phenomenex Standard Terms and Conditions which may be viewed at www.phenomenex.com/TermsAndConditions.

Trademarks

Rezex, SecurityGuard, and Phenex are trademarks of Phenomenex. Agilent is a registered trademark of Agilent Technologies, Inc. arium[®] is a registered trademark of Sartorius AG.

Disclaimer

Dimensions and chromatographic conditions are the same for all columns unless otherwise noted.

SecurityGuard is patented by Phenomenex, U.S. Patent No. 6,162,362

CAUTION: this patent only applies to the analytical-sized guard cartridge holder, and does not apply to SemiPrep, PREP or ULTRA holders, or to any cartridges.

© 2015 Phenomenex, Inc. All rights reserved.