

# APPLICATIONS

# Extraction and Analysis of Vitamin B1 (Thiamine) from Guar Gum by HPLC-UV

## Scott Krepich and Morgan Kramer

Phenomenex, Inc., 411 Madrid Ave., Torrance, CA 90501 USA



Scott Krepich Senior Field Application Scientist Scott enjoys surfing and eating. He is crazy about chromatography, because his mom is really into CSI

and thinks that is what he does.

in 🎔

# Introduction

Guar Gum is a natural substance made from guar beans, and is commonly used in the food industry as an emulsifier, stabilizer, and thickener. Quantifying components formulated in such can be a challenge due to the matrix viscosity and potentially interfering components.

Here we demonstrate an effective extraction for HPLC-UV quantification of Vitamin B1 in matrix, that may be expanded to cover additional water soluble vitamins.

# **LC Method Parameters**

Column:	Luna <sup>®</sup> Omega	3 µm Polar C18
Dimensions:	150 x 4.6 mm	
Part No.:	00F-4760-E0	
Mobile Phase:	A: 20 mM Soc	lium phosphate, pH 3.5 with phosphoric acid
	B: Methanol	
Gradient:	Time (Min)	%B
	0	0
	15	60
	15.1	0
Flow Rate:	0.5 mL/min	
Injection Vol.:	1 µL	
Temperature:	22°C	
Detector:	UV @ 254 nm	

# **Sample Preparation**

- 1. Weigh out ~2g of guar gum into a 125mL Erlenmeyer flask
- Add between 30-50 mL of the extraction solvent (see Table 1 for trial matrix) MeOH-2 (Methanol : 1 % HCL in water 80:20)
- Shake vigorously until all guar gum appears to have absorbed/ dissolved into the solvent
- 4. Sonicate for 10 minutes
- 5. Pass an aliquot (~2 mL) of the mixture through a PTFE syringe filter
- 6. Inject  $1 \,\mu L$  of the filtrate using the HPLC method shown above

## Table 1.

**Extraction Solvent Trials** 

Extraction Labels	Solvents	Solvent Ratio	Volume added (mL)
MeOH	Methanol:1 % HCl in water	80:20	50
EtOH – Recrystallization*	1 % HCl in water	100	35
ACN H+	Acetonitrile + 0.1 % Acetic Acid	100	45
ACN -2	Acetonitrile: 1 % HCl in water	80:20	50
IPA	lsopropanol: 1 % HCl in water	80:20	50
EtOH -2	Ethanol: 1 % HCl in water	80:20	50
NaCl	5 % NaCl dissolved in 1 % HCl	100	150
Me0H -2*	Methanol: 1 % HCl in water	80:20	50
Acetone	Acetone: 1 % HCI in water	80:20	50
Water	Water	100	50
H+, H <sub>2</sub> 0	1 % HCl in water	100	50

\* The EtOH and MeOH-2 –Recrystallization had an additional step where 5 mL of the dissolved guar gum sample was aliquoted into a test tube and cooled in an ice bath. Followed by the addition of 5 mL cooled ethanol to induce crystalization of undesired material.



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Note: Peak areas obtained by employing the above "Running Conditions" and relating peak areas obtained to concentration of known standards

# Table 2.

Quantitation of Thiamine Recovered from Extraction

Extraction from A1973-VS5	Guar Powder (g)	Peak Area (B1)	Mass Thiamine (mg)	Projected Conc. (mg/lb)**	ldeal conc. Range (mg/lb)
Extractions Techniques					
MeOH	2.36	61.4	0.78	149.62	100-200
EtOH – Recrystallization	2.62	17.0	0.30	51.42	100-200
ACN H+	2.29	10.1	0.11	22.13	100-200
ACN -2	2.14	11.8	0.01	3.09	100-200
IPA	2.30	46.9	0.59	117.05	100-200
EtOH -2	1.94	14.3	0.18	41.55	100-200
NaCl	2.14	16.7	0.62	132.46	100-200
MeOH -2	3.11	87.6	1.11	162.49	100-200
Acetone	2.42	19.3	0.24	45.33	100-200
Water	2.42	59.8	0.76	141.85	100-200
H+, H <sub>2</sub> 0	2.43	80.5	1.02	190.66	100-200

Note: Quantitation was performed by relating peak area to concentration from the developed HPLC method and calibration curve (Figure 1).

\*\*Projected Conc. was calculated assuming each 100% recovery from each extraction. Values are reported as mg of thiamine per pound of guar powder.

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mAU 60 -40 -20 -0 -

10

20 min

# **Results and Discussion**

\* The arrow identifies the Thiamine peak

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Developed is a rapid, simple, and robust HPLC method for the separation and retention of Thiamine from a guar gum sample. The novel polar functionality within the Luna Omega Polar C18 stationary phase help improve retention and peak shape without the need for ion-pairing agents in the mobile phase.

The extraction procedures developed shone light onto which methodologies were most efficient in the extraction of thiamine from the matrix. In order to maintain a direct comparison among extraction techniques, only the guar gum sample labeled A 1973-VS5 was used for this analysis. The provided method, using and 80:20 mixture of Methanol to 1 % HCl was used as the comparison point for subsequent extractions. In **Figure 2**, the MeOH and MeOH-2 extractions are representative of the method being used by the client. These extractions gave results that led to a calculation of 149.62 and 162.49 µg/lb respectively for the guar gum sample. These results are consistent with the predicted range of 100-200 µg/lb.

Most of the extractions netted a lower yield than the MeOH and MeOH-2 extractions, though the H+, H<sub>2</sub>O extraction managed to have a higher recovery than the MeOH extractions. The H+, H<sub>2</sub>O

Figure 3.







extraction yielded results that allowed for the calculation of thiamine to be 190.66  $\mu$ g/lb. This value is consistent with the predicted range, though, the higher thiamine concentration being calculated through this extraction indicates a greater recovery from this extraction technique. The calculated projected concentration of thiamine in the guar gum sample was done in order to compare the extraction techniques. A higher projected thiamine concentration as a result of the extraction, in turn, means a higher percentage of thiamine was recovered from the guar gum. The projected concentration was in order to compare the results gathered from the assay to the expected values provided by the client and as a way to monitor successful extractions.

Each of the extractions were done in duplicate, in order to ensure consistency in the results. Further work must be done in order to fully represent percent recovery, account for matrix effects, develop RSDs for the extractions, and to determine differences in recovery for the various samples. In conclusion, we provide a fast, robust, and ion-pairing agent free HPLC separation and quantitative assay for Thiamine. Additionally, we have laid the groundwork for further optimization of Thiamine extraction from the guar gum matrix.

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# Luna Omega<sup>™</sup> Ordering Information

1.6 µm Microbore and Minibore Columns (mm)			SecurityGuard ULTRA Cartridges (mm)				SecurityGuard A Cartridges (mm)	
Phases	50 x 1.0	100 x 1.0	150 x 1.0	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	3/pk
Polar C18	00B-4748-A0	00D-4748-A0	00F-4748-A0	00A-4748-AN	00B-4748-AN	00D-4748-AN	00F-4748-AN	AJ0-9505 for 2.1 mm ID
3µm Minibor	e and MidBore™ Col	umns (mm)						SecurityGuard Cartridges (mm)
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	50 x 3.0	100 x 3.0	150 x 3.0	4 x 2.0*
Polar C18	00A-4760-AN	00B-4760-AN	00D-4760-AN	00F-4760-AN	00B-4760-Y0	00D-4760-Y0	00F-4760-Y0 for ID:	AJ0-7600 2.0 - 3.0 mm
3µm Analytic	al Columns (mm)				SecurityGuard Cartridges (mm)			
Phases	50 x 4.6	100 x 4.6	150 x 4.6	250 x 4.6	4 x 3.0*			
Polar C18	00B-4760-E0	00D-4760-E0	00F-4760-E0	00G-4760-E0	AJ0-7601			
				for ID:	3.2-8.0 mm			
5µm Minibor	e Columns (mm)				SecurityGuard Cartridges (mm)			
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	4 x 2.0*			
Polar C18	00A-4754-AN	00B-4754-AN	00D-4754-AN	00F-4754-AN	AJ0-7600			
				for ID:	2.0 - 3.0 mm			
5um MidBore	e™ Columns (mm)			SecurityGuard Cartridges (mm)				
Phases	50 x 3.0	100 x 3.0	150 x 3.0	4 x 2.0*		lauai	ranteel	
Polar C18	00B-4754-Y0	00D-4754-Y0	00F-4754-Y0	AJ0-7600		90.0.		
			for ID:	2.0 - 3.0 mm		If Luna a	nalutioal oolumn	a da nat provida at laga
5µm Analytic	al Columns (mm)				SecurityGuard Cartridges (mm)	equivale	nalytical column nt separation as	s compared to a compe
Phases	50 x 4.6	100 x 4.6	150 x 4.6	250 x 4.6	4 x 3.0*	dimensio	ons return the	column with compara
Polar C18	00B-4754-E0	00D-4754-E0	00F-4754-E0	00G-4754-E0	AJ0-7601	data with	nin 45 days for a	
				for ID:	3.2-8.0 mm			CI OLLI ILI OND.
* SecurityGuar	d ULTRA Cartridges rea	quire holder, Part No.:	AJ0-9000			Terms and	Conditions	

SecurityGuard ULTRA Cartridges require holder, Part No.: AJ0-9000 SecurityGuard Analytical Cartridges require holder, Part No.: KJ0-4282

### Australia

- t: +61 (0)2-9428-6444 f: +61 (0)2-9428-6445
- auinfo@phenomenex.com

### Austria

- t: +43 (0)1-319-1301 f: +43 (0)1-319-1300
- anfrage@phenomenex.com

- **Belgium** t: +32 (0)2 503 4015 (French) t: +32 (0)2 511 8666 (Dutch)
- f: +31 (0)30-2383749
- beinfo@phenomenex.com

## Canada

- t: +1 (800) 543-3681 f: +1 (310) 328-7768
- info@phenomenex.com

# China

t: +86 400-606-8099 f: +86 (0)22 2532-1033 phen@agela.com

- Denmark
- t: +45 4824 8048 f: +45 4810 6265 nordicinfo@phenomenex.com

# www.phenomenex.com

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- Finland t: +358 (0)9 4789 0063
- f: +45 4810 6265
- nordicinfo@phenomenex.com

# France t: +33 (0)1 30 09 21 10

- f: +33 (0)1 30 09 21 11
- franceinfo@phenomenex.com

# **Germany** t: +49 (0)6021-58830-0

- f: +49 (0)6021-58830-11
- anfrage@phenomenex.com

## India

- t: +91 (0)40-3012 2400 f: +91 (0)40-3012 2411
- indiainfo@phenomenex.com
- Ireland
- t: +353 (0)1 247 5405 f: +44 1625-501796
- eireinfo@phenomenex.com
- Italy t: +39 051 6327511

# f: +39 051 6327555

italiainfo@phenomenex.com

# **Norway** t: +47 810 02 005

# Puerto Rico

# t: +1 (800) 541-HPLC

# Spain

- t: +34 91-413-8613 f: +34 91-413-2290
- espinfo@phenomenex.com

### Sweden

- t: +46 (0)8 611 6950 f: +45 4810 6265
- nordicinfo@phenomenex.com

### United Kingdom

- t: +44 (0)1625-501367
- f: +44 (0)1625-501796 ukinfo@phenomenex.com

### USA

- t: +1 (310) 212-0555 f: +1 (310) 328-7768
- info@phenomenex.com

# All other countries Corporate Office USA

- f: +1 (310) 328-7768
  - info@phenomenex.com
- TN53180617\_W

# **Luxembourg** t: +31 (0)30-2418700 f: +31 (0)30-2383749 nlinfo@phenomenex.com

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- **Mexico** t: 01-800-844-5226 f: 001-310-328-7768
- tecnicomx@phenomenex.com

# The Netherlands

- t: +31 (0)30-2418700 f: +31 (0)30-2383749 nlinfo@phenomenex.com

### New Zealand

- t: +64 (0)9-4780951 f: +64 (0)9-4780952
- nzinfo@phenomenex.com

- f: +45 4810 6265 nordicinfo@phenomenex.com

- f: +1 (310) 328-7768
  - info@phenomenex.com

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